

**In the Specification**

Starting on page 21, please amend paragraph [0084] as follows:

[0084] The total RNAs were produced by the same technique as described above. The quality and cleanliness of the RNAs were verified on denaturing gel. The reverse transcription was performed using the Qiagen Omniscript™ kit in a specific manner with a 20 mers LTR primer. The conditions used were 10 ng of LTR primer, 2.5 mM of dNTP, 1 µg of total RNA supplemented by 2 OU of RNase inhibitor (RNAsin®), buffer RT and 4 OU of reverse transcriptase. The mixture was incubated for one hour at 37°C and for 5 minutes at 94°C. The RT products of the specific cDNAs were amplified by PCR using two other primers named as 1 and as 2. The reaction conditions were 0.25 mM dNTP, 100 ng of each of the primers and 1/20<sup>th</sup> of the RT product, 1.5 mM MgCl<sub>2</sub>, buffer Taq pol (Perkins Elmer N801-0060). The cycles employed were 4 minutes 94°C, 45 s 61°C, 1 min 72°C) and 10 minutes at 72°C.

LTR: AGATATCCTGTTGGCCAT                           SEQ ID NO:1

AS1: GCCGTGCATCATCCTGACTG                           SEQ ID NO:2

AS2: CTGTTCCTGACCTTGATCTG                           SEQ ID NO:3

Tumorigenicity test

On page 22, please amend paragraph [0086] as follows:

[0086] The cDNA expression card “Atlas cDNA expression Arrays” (Clontech) was implemented according to the supplier’s instructions. Two identical nylon membranes (no. 7741-1) on which were deposited 588 samples of murine cDNA, corresponding to 588 genes, made possible parallel hybridization of the cDNAs of two different cell lines. Information regarding the 588 genes

is available [[on]] in the Clontech web site (<http://www.clontech.com/atlas.genelist/search.htm>) catalogue. The preparation of the radiolabeled cDNA probes was implemented by reverse transcription with [<sup>32</sup>P] dATP using the Clontech kit. Hybridization was performed according to the supplier's instructions. The signals were observed with the PhosphoImager.